

INTRODUCTION

Solar salterns consist of a series of shallow ponds connected in a sequence of increasingly saline brines. Crystallizers are the last ponds which have a salinity above 30%. Archaea (and to a lesser extent Bacteria) are common in communities associated with hypersaline environments. Viruses are also present in these ecosystems and they have been estimated to occur in the range of 10⁹ virus-like particles (VLP) per ml. (Santos *et al.*, 2012). Interestingly, the number of viruses, which is normally correlated to the number of cells, increases with the salt concentration. The importance of viruses in ecosystems lies in the regulation of microbial community composition through highly specific host-virus interactions (Weinbauer & Rassoulzadegan, 2004). Although most of the abundant extremely halophilic Archaea and Bacteria can be cultivated, no viruses were isolated so far that infect them. (Santos *et al.*, 2012).

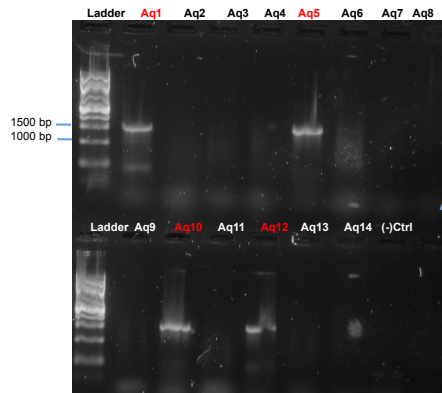
Hypothesis

There must be viruses in hypersaline environments that can infect and lyse Bacteria/Archaea isolated from the same location.

Objective

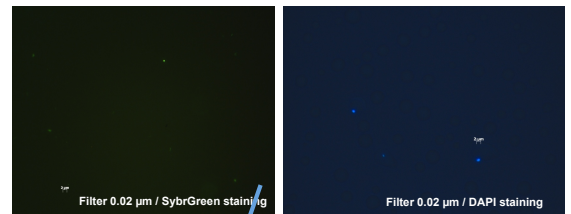
Establish the presence of viruses in a solar saltern using a culture-dependent approach with Bacteria/Archaea hosts and demonstrate lysis under laboratory conditions.

2. PCR-based detection of 16S rRNA genes



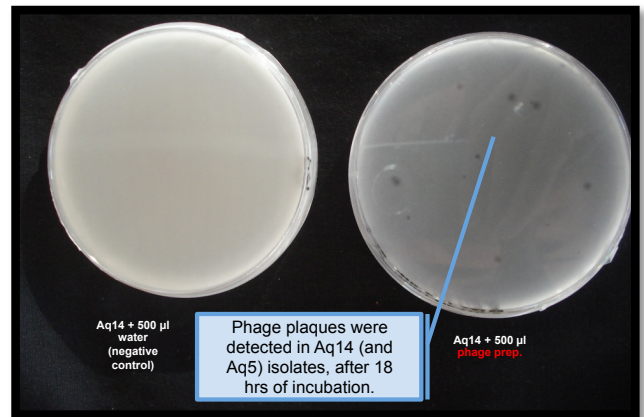
Aq1, Aq5, Aq10 and Aq12 isolates were positive for Bacteria 16S rRNA gene. The remaining isolates might be Archaea strains.

3. Epifluorescence microscopy of cell-free-concentrated filtrates



A few putative virus-like particles were observed.

4. Infection Assay



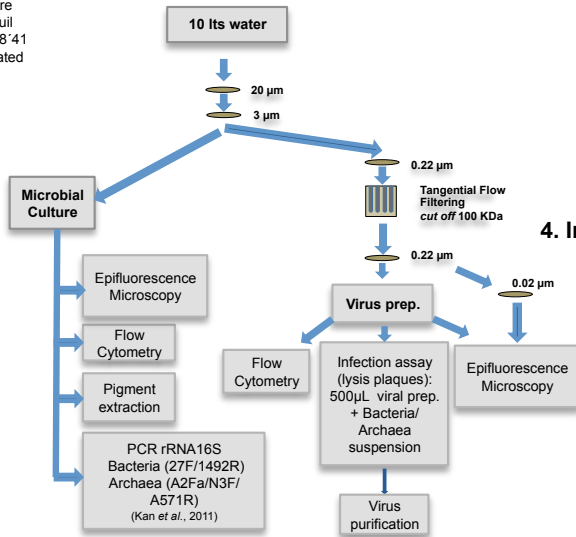
Phage plaques were detected in Aq14 (and Aq5) isolates, after 18 hrs of incubation.

SAMPLING AREA



Water samples were collected at the Cahuil solar salterns (34° 28' 41" S / 72° 1' 6" W), situated 14 km south of Pichilemu (region O' Higgins, Chile)

METHODOLOGY

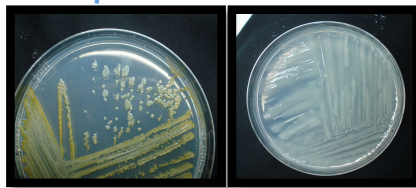


RESULTS

1. Bacteria/Archaea Isolation

NaCl % in medium	Carbon Source	N° of Isolates	% Isolates
3,5 %	peptone	12	66.67
7,5%	peptone	2	11.11
12%	peptone	1	5.56
18%	peptone	1	5.56
18%	glucose	1	5.56
18%	glycerol	1	5.56
TOTAL		18	100

After 48-72 hrs of incubation on solid medium, most colonies grew in low (3.5%) NaCl concentration. Some strains developed orange/pink colours.



CONCLUSIONS / FINAL REMARKS / PERSPECTIVES

- A morphologically diverse collection of Bacteria/Archaea was obtained in low NaCl-containing solid media. Isolation of more extreme halophiles would require extended (several weeks) incubation periods.
- As expected for this extreme environment, bacterial 16S rRNA gene was amplified only in a minority of strains. Detection of archaeal 16S rRNA gene would provide a more complete and reliable picture of the community composition.
- Viral lytic infection (plaque development) in Aq5 (Bacteria) and Aq14 (no Bacteria, possibly Archaea) strains was observed. Since we detected different plaque morphologies (clear vs. turbid-bordered plaques) we could infer that more than one viral strain lysed the hosts.

BIBLIOGRAPHY:

- Dyall-Smith M. The Haloarchaeal Handbook: Protocols for Halobacterial Genetics. 2008. p. 144. <http://www.haloarchaea.com>
- Santos, F., P. Yarza, V. Parro, I. Meseguer, R. Rosselló-Móra, and Josefa Antón et al 2012. viruses from hypersaline environments: a culture-independent approach. Appl. Environ. Microbiol. AEM.07175-11; published ahead of print 13 January 2012, doi:10.1128/AEM.07175-11
- Weinbauer, M. G. and Rassoulzadegan, F. 2004. Are viruses driving microbial diversification and diversity? Environ. Microbiol. 6: 1–11

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